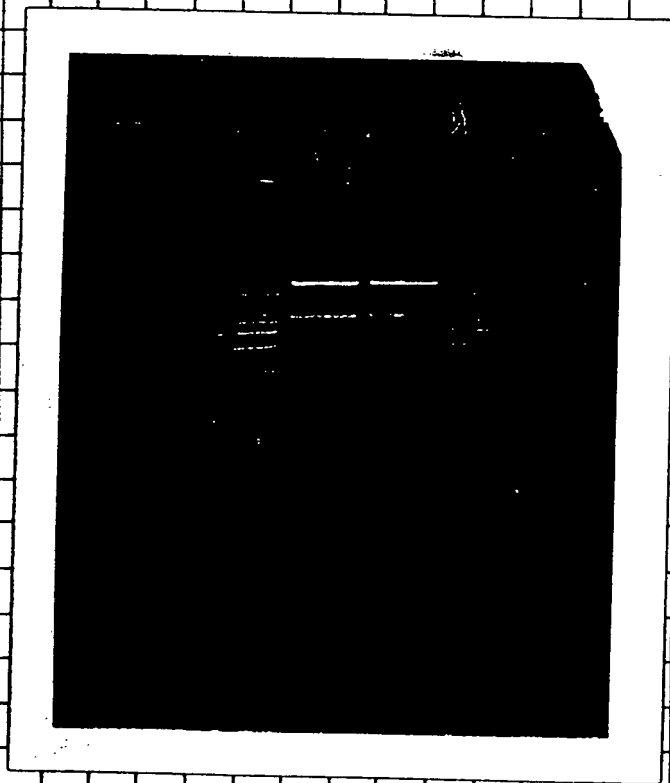


n Page No. 9

EcoRI / Bam HI RDS on 1 Hep 5+6
ex rxn mixed

10µl 1 DNA
5µl 10x "B"
1µl EcoRI
1µl Bam HI
33µl H₂O
50

Inc 37°C ~ 2 hrs → added 10µl 5x dye to each
2µl each on 0.7% agarose (1x TBE)



Set up a 1% CMP gel
for preparative
electrophoresis on 2/20.
Apparently 15+6 at
least have the
full length coding
regions for Tyro10.

To Page No. 11

Witnessed & Understood by me,

Date

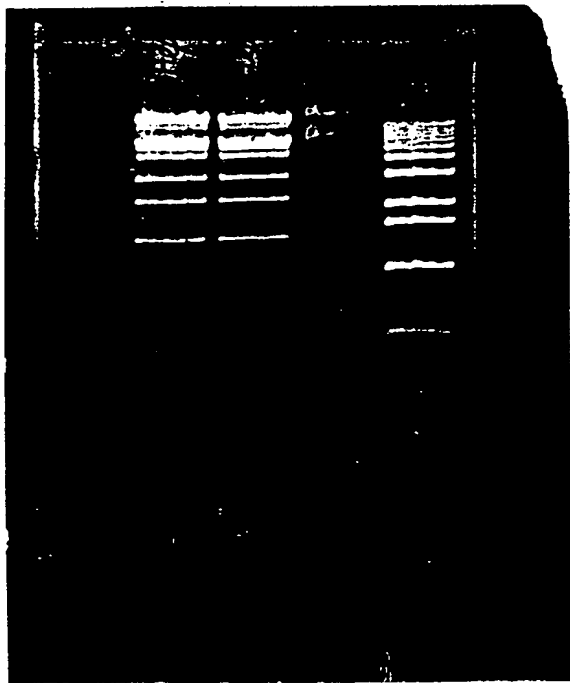
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From Page No. 10

Ran remaining 50µl 1546 RD's on 1% agarose
cut out indicated bands



Ran Magic PCR preps
collected each in 50µl
combined

Ligation to pRK5
mixed for each

1µl pRK5 ÷ EcoRI/BamHI / ~~0.001~~
2µl 10x ligase buffer
1µl Ligase
7µl insert (or 1µl)
8µl H₂O (or 6µl)

Plus a vector alone ligation
= 3 total.

Inc 12.5°C O/N.

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transformed O/N pK5 Ligs to competent
XL-2 blue cells.

Plated each onto 5 x 100mm LB cant⁵⁰ plates
(cont. plated onto 2 plate)
Inc all (11) 37°C O/N.

To Page No. 13

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WIM Bacon

TITLE

Book No. 175

Exhibit B, pg. 4 of 7

From Page No. 12

RD of SKG / CH₂CH₃ IgG1 fusion vector

mixed

2 µl DNA (~4 µg)
 5 µl 10x "B"
 4 µl H₂O
 1 µl EcoRI
 1 µl BstEII
 50

Inc 37°C ~ 2 hrs

Ran on 0.7% agarose

Isolated band on N445 paper
 Eluted in 400 µl 1M NaCl (in TE) 70°C 2.
 Removed paper.
 Stored - 70°C o/n.

Ordered PCR primers to PCR ECD of HPI
 & incorporate a BstEII site @ 3'e

Genentech, Inc.
 Genentech, Inc.

SYNTHETIC DNA REQUEST A-6749 ✓

PROJECTOR WILL BREON EXTENSION 2650 LAB NO. 10231 PROJECT NAME OR NUMBER 1713 DATE NEEDED AMT NEEDED

LIST SEQUENCE(S) & 3' NOTE AS SUCH

SIZE & FRAGMENT NAME:

PLEASE INDICATE BY "*" FRAGMENT(S) TO BE CLONED

1x 24mer
 1x 35mer

① Tyro P13 (24mer)
 (5') TGG·GAG·GAG·GAG·CCC·ATG·CGC·CAC·3' "

② Tyro P14 (35mer)
 (5') GTA·CAG·TNA·CCG·GCG·GTC·GAG·CTC·
 ·CCC·TCG·GAC·TT·3' "

FRAGMENT USE: ☐ PRIMER ☐ PROBE ☒ PCR ☐ GENE CONSTRUCTION ☐ LINKER/ADAPTOR

☐ MUTAGENESIS ☐ OTHER (SPECIFY):

SPECIAL REQUESTS

WILL BREON DATE APPROVED PAPER SIGNATURE DATE

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To Page N

om Page No. 13

Checked PRK5/HPTK6 (tyro10) transformation plates
Got 39 transformants

Started 39 x 5ml 1B cult⁵⁰ (+ master plate)

Inc all 37°C O/N.

To Page No. 1

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Date

Invented by

11/7/13 Balman

Date

11/7/13

From Page No. 14

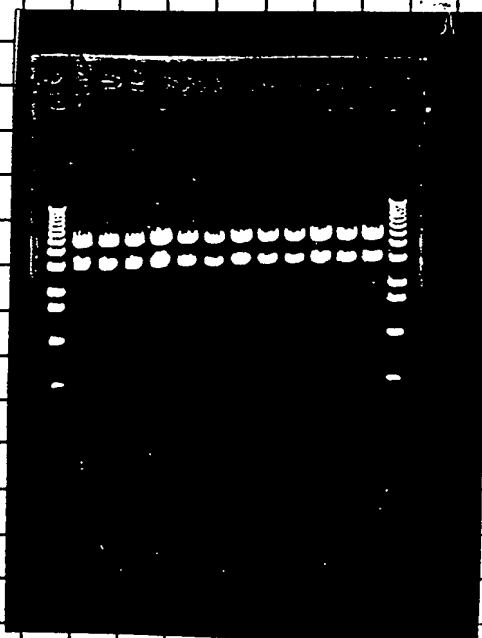
Did Magic MP's on 12 of Q/N PRK5/HPTK6 MP's
(Stored remaining 27 cultures + master @ 9

Collected each in 50 μ l TE

RD's
Per rxn 2 μ l MP DNA
 2 μ l 10x "B"
 15 μ l H₂O
 0.5 μ l EcoRI
 0.5 μ l BamHI
 20

Inc 37°C ~ 1.5 hrs \rightarrow added 4 μ l dye to each

Ran ~ 15 μ l each on 0.7% agarose (1x TBE)



All positives.

Started 1x 500 μ l ZYT carb⁵⁰ MIDI
prep on #1

Inc 37°C O/N.

To Page

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Date

Invented by

H. H. Benson

Date

m Page No. 15

torced 1.2ml O/N HPTK6/pRK5 + 400µl 50% glycerol
E70°C

Did Magic Maxi prep on remainder of culture

EtoH ppt'd DNA after elution from resin column

Stored ppt'd pellet in -20°C 70% EtoH. O/N

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Date

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Recorded by

Date

WIM Bacon